### **SUPPLEMENTARY DATA**

## SUPPLEMENTARY METHODS

### **Echocardiographic studies**

Transthoracic echocardiograms were performed in all patients with General Electric Vivid E9 equipment (GE Healthcare Vingmed, Trodheim) as part of the work-up to exclude potential causes of stroke. All images were stored for subsequent off-line analysis in the EchoPAC v201 workstation. Echocardiographic measurements were acquired and analyzed following international recommendations.<sup>1</sup> Echocardiogaphic evaluation included measurement of standard parameters regarding LV dimensions and function, and, specifically, anatomical and functional assessment of the left atrium (LA), which included evaluation of LA anteroposterior diameter, LA biplanar area, LA biplanar volume (calculated from the 4-chamber and 2-chamber apical axes using the area-length method), and left atrial ejection fraction according to the formula: maximum LA volume – minimum LA volume / maximum LA volume x 100.<sup>1</sup>

### Insertable cardiac monitor implantation and atrial fibrillation definition

Insertable cardiac monitor devices (Biomonitor and Biomonitor-2, Biotronik) were implanted subcutaneously under local anesthesia in the left chest region and were programmed with the specific algorithm for AF detection, defined by an RR interval variability > 12.5% lasting > 1 minute, as previously described.<sup>2</sup> RR intervals with cycle lengths below 200 ms were considered noise. All patients were included in a remote monitoring system (Biotronik Home Monitoring). Recordings automatically detected as potential AF episodes were reviewed by a specialized cardiologist. AF was defined by the presence of a confirmatory revised insertable cardiac monitor recording.

## MicroRNA extraction and profiling

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MiRNAs were isolated from plasma using the miRNeasy Serum/Plasma kit (Qiagen), and retrotranscribed into cDNA with the TaqMan Advanced miRNA cDNA Synthesis Kit (Life Technologies). In the discovery phase, a TaqMan OpenArray Human Advanced MicroRNA Panel (ThermoFisher) was carried out to screen 754 of the most commonly expressed human miRNAs. Briefly, cDNA was mixed 1:80 with the TaqMan master mix and cycled on a QuantStudio 12K Flex. Expression levels of all miRNAs were analyzed with the Cloud software (ThermoFisher), specifically designed for high-throughput gene expression studies. Mean global normalization was used to normalize samples. Those miRNAs found to have a significant differential expression among groups (P < .05 and fold change > 2 or < 0.5) and/or were considered of potential relevance with regards to AF were selected for replication. MiRNA determination in the replication phase was performed by qPCR using the Fast Advanced master mix (ThermoFisher) with specific TaqMan miRNA probes (Life Technologies), and cycled in a 7500 ThermoCycler (Applied Biosystems). Results were analyzed using the Expression Suite software (Life Technologies). MiR-16, miR-92a and miR-486 showed high stability between groups and were therefore used as controls for normalization.

	Set I (discovery phase, matched for age and sex)			Set II (replication phase, unmatched)		
	CrS-SR (n =	CrS-AF (n = 9)	CES-AF (n =	CrS-SR (n = 27)	CrS-AF (n = 19)	Р
	9)		9)			
Age, y	75.6 ± 8.2	75.2 ± 7.6	74.7 ± 8.0	72.2 ± 12.8	77.4 ± 10.0	.145
Female sex	4	4	4	14 (51.9)	7 (36.8)	.377
BMI, kg/m <sup>2</sup>	28.1 ± 3.3	29.6 ±6.4	27.8 ±3.7	27.1 ± 4.1	26.4 ± 4.5	.624
Hypertension	7	5	7	21 (77.8)	12 (63.2)	.331
Diabetes mellitus	2	1	3	7 (25.9)	3 (15.8)	.488
Hyperlipidemia	5	5	5	16 (59.3)	9 (47.4)	.550
Current smoking	1	1	2	4 (14.8)	2 (10.5)	.999
Peripheral artery	1	2	1	3 (11.1)	5 (26.3)	.246
disease						
Coronary artery disease	0	0	1	1 (3.7)	2 (10.5)	.561
Prior stroke/TIA	2	3	1	3 (11.1)	3 (15.8)	.680
CHA <sub>2</sub> DS <sub>2</sub> -VASc	3.3 ± 2.2	3.3 ± 1.9	3.4 ± 1.3	3.1 ± 1.6	3.1 ± 1.9	.991
TIA/stroke*	1/8	0/9	1/8	6 (22.2)	4 (21.1)	.999

## Table 1 of the supplementary data. Characteristics of Set I and Set II populations

AR, atrial fibrillation; BMI, body mass index; CES, cardioembolic stroke; CrS, cryptogenic stroke; SD,

standard deviation; SR, sinus rhythm; TIA, transient ischemic attack.

The data are expressed as absolute number, No. (%), or mean ± standard deviation.

\* In population Set II, the numbers of TIA are presented.

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# Table 2 of the supplementary data. Results of the open array study

Differential miRNA expression CrS-AF vs CrS-SR			Differential miRNA expression CES-AF vs CrS-SR				
miRNA	Rq	Р	miRNA	Rq	Р		
hsa-miR-320a*	362.020	<.001	hsa-miR-320a*	344.390	<.001		
hsa-miR-362-5p*	0.388	.005	hsa-miR-19b-3p	0.017	.003		
hsa-miR-744-5p	10.594	.007	hsa-miR-190a-	2.889	.003		
			5p*				
hsa-miR-664a-	0.389	.009	hsa-miR-362-5p*	0.330	.005		
3p*							
hsa-miR-1-3p	8.644	.014	hsa-miR-744-5p	10.547	.011		
hsa-miR-377-3p	4.136	.015	hsa-miR-214-3p*	0.418	.013		
hsa-miR-181b-5p	4.686	.017	hsa-miR-181d-5p	4.514	.014		
hsa-miR-190a-	2.227	.021	hsa-miR-1180-	0.395	.017		
5p*			3p*				
hsa-miR-19a-3p	0.152	.022	hsa-miR-1-3p	7.932	.018		
hsa-miR-483-5p*	2.439	.024	hsa-miR-31-5p*	0.411	.019		
hsa-miR-425-5p	3.262	.026	hsa-miR-34a-3p*	0.411	.019		
hsa-miR-16-2-3p*	0.324	.028	hsa-miR-455-3p*	0.411	.019		
hsa-miR-625-3p*	0.413	.031	hsa-miR-499a-	0.411	.019		
			5p*				
hsa-miR-502-3p*	1.862	.031	hsa-miR-570-3p*	0.411	.019		
hsa-miR-191-5p	4.758	.039	hsa-miR-29b-2- 5p*	0.435	.021		
hsa-miR-19b-3p	0.128	.049	hsa-miR-154-3p*	0.434	.021		
			hsa-miR-548e-	0.458	.022		
			3p*				
			hsa-miR-548j-5p*	0.454	.024		
			hsa-miR-100-5p*	0.500	.025		
			hsa-miR-188-5p*	0.465	.025		
			hsa-miR-452-5p*	0.453	.025		
			hsa-miR-23b-5p*	0.461	.027		
			hsa-miR-493-3p*	0.518	.029		
			hsa-miR-17-5p	6.432	.031		
			hsa-miR-99b-5p	4.144	.032		
			hsa-miR-500a- 5p*	0.479	.036		
			hsa-miR-425-5p	2.970	.037		
			hsa-miR-29c-3p	13.762	.037		
			hsa-miR-380-3p*	2.070	.038		
			hsa-miR-941*	0.420	.038		
			hsa-miR-532-3p	2.780	.039		
			hsa-miR-186-5p	4.605	.039		
		-	hsa-miR-377-3p	4.277	.041		

AR, atrial fibrillation; CES, cardioembolic stroke; CrS, cryptogenic stroke; SR, sinus rhythm.

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\*Inconsistent values of Ct among samples (high standard deviation), or very low (Ct > 36)

expression in both groups. miRNAs selected for replication are highlighted in bold.